



Research article

Isolation, characterization and anti-inflammatory activity of compounds from the *Vernonia amygdalina*

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ABSTRACT

The need to explore the abundance of natural products cannot be overemphasized particularly in the management of various disease conditions. In traditional medical practice, *Vernonia amygdalina* has been widely adopted in the management of various inflammatory disorders. The objective of this investigation was to isolate the bioactive principles from the stem-bark and root of *V. amygdalina* and assess the anti-inflammatory (*in vitro*) activity of both the crude extracts and the isolated compounds. Following extraction with the methanol, the extract was subjected to gravity column chromatography and the resultant fractions was further purified to obtained pure compounds. The structural elucidation of the compounds were based on data obtained from ¹H to ¹³C nuclear magnetic resonance (NMR) spectroscopies as well as fourier transform infrared (FT-IR). Using diclofenac as a control drug, the albumin denaturation assay was used to determine the *in vitro* anti-inflammatory activity of the extracts and isolates. Three distinct compounds characterized are vernoamyoside D, luteolin-7- α -o-glucuronide, and vernotolaside, a new glycoside. When compared to diclofenac, which has an IC₅₀ of 167.8 μ g/mL, luteolin-7- α -o-glucuronide, vernoamyoside D, and vernotolaside all showed significant inhibitions with respective IC₅₀ values 549.8, 379.5, and 201.7 μ g/mL. Vernotolaside is reported for the first time from the root. The assertion that the plant is used in traditional medicine for the management of inflammatory disorder is somewhat validated by the confirmation of the existence of the compounds with the

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biochemical actions. Further validation of the isolated compounds would be required in animal studies.

1. Introduction

Vernonia amygdalina of the family Asteraceae is one of the most nutritional culinary medicinal plants in primarily in the tropical regions. On the account of his bitter taste, it is often referred to as “bitter leaf” [1]. *V. amygdalina* is a perennial shrub that reaches 3 m in height in the tropics [2]. The plant is grown by some tribes who uses it for culinary purpose in Nigeria [3]. The macerated leaves of the plant are consumed as vegetables and condiments. Interestingly, the aqueous leaf extract is applied topically for wound healing [4, 5]. The plant is renowned for its protective effects in various genotoxicity, cardiotoxicity and hepatotoxicity studies [6–8]. The crude extract of the leaf is also reported to possess anti-inflammatory potential [9–11] and synergistic anti-cancer effects with gemcitabine [12]. The plant have many other non-medicinal applications particularly in nanoparticle preparations [13]. Bioactive principles have been obtained from various parts of plants which include the leaves, stem, wood, root, flowers and seed [14]. The root of plant has been a source isolation of bioactive principles [15–18]. While nanoparticles have been used for various things like biosensor preparation [19,20], the nanoparticles produced from the plant exhibited anti-inflammatory and antinociceptive activities in the animal model [21,22]. In this study, we isolated and characterized the major bioactive principles from *V. amygdalina* root and stem bark. The *in vitro* anti-inflammatory potentials using albumin denaturing inhibition assay was also evaluated.

2. Materials and Methods

2.1. General

A silica gel 60F₂₆₄ Merck, Darmstadt, Germany precoated TLC plates (0.2 mm layer thickness) was used for spotting and visualisation carried out under short and long wavelengths of the UV-lamp or by reaction of the developed plate with iodine vapour in the iodine chamber for 1 min. Fractionation was performed on silica gel-loaded column chromatography (CC). For the determination of the functional groups in the isolated compound, a Shimadzu (8400s) Fourier Transform-Infrared (FT-IR), Shimadzu, Japan, was used to generate the infrared spectra using potassium bromide pellets. ¹H and ¹³C NMR spectra were obtained in chloroform-*d*₃ on Bruker 400 Nuclear Magnetic Resonance Spectrometer operating at 400 MHz for proton and 75 MHz for carbon. Chemical shifts were recorded in ppm downfield of tetramethylsilane (TMS).

2.2. Preparation of Plant materials

The plant material (stem-bark and root) of *V. amygdalina* were harvested fresh from a botanical farm in Ilorin and authenticated by renowned taxonomist at the herbarium of the department of plant biology, University of Ilorin, Ilorin, Nigeria where the voucher (specimen) number UIL/001/972/2021 was documented.

2.3. Extraction, column chromatography Fractionation and isolation

The roots and stems were carefully rinsed with water to remove dirt and attached soil. The plant materials were shredded dried at ambient temperature without exposure to direct sunlight. They were blended into fine powder using electrical blender. The blended plant materials were extracted successively with solvents of increasing polarities separately. The mixtures was filtered and concentrated on a rotary evaporator (BUCHI) at reduced pressure. The extracts were stored in a refrigerator for further use. The methanol root and stem-bark extracts were dissolved in small amount of methanol and chromatographed independently over silica gel-loaded column using n-hexane, ethyl-acetate and methanol as eluting solvent system in polarity gradient order. 80 and 90 fractions were collected for the methanol root extract and methanol stem-bark extract respectively. Following TLC profiling, the fractions were combined to give 5 and 9 subfractions respectively. The subfractions obtained were subjected to another chromatographic elutions in a silica gel loaded column and some partially purified compounds that resulted were further purified via preparative thin layer chromatography or washing with selected solvents. Spectroscopic elucidation and comparison with literature data afforded three compounds which include luteolin-7- α -o-glucuronide (a flavonoid glycoside), vernoamyoside D (a triterpene glycoside), and vernotolaside (a bifuran glycoside).

2.4. In vitro albumin denaturing inhibition assay

The *in vitro* albumin denaturing inhibition assay was determined following standard protocol [23,24]. In order to obtain 2 mL reaction mixture, 1 mL 20 mM Tris HCl and 0.06 mg trypsin, buffered at pH 7.4 with the test samples (1 mL) of dilution range, 5–250 μ g/mL was thoroughly mixed and incubated at 37 °C for 5 min before addition of 1 mL of prepared 0.8 % casein. The resultant mixture was inhibited for additional 20 min before 2 mL 70 % perchloric acid was adopted to terminate the reaction progress. The colloidal mixture that resulted was subjected to centrifugation and the absorbance taken at 210 nm while the buffer was adopted as blank. For each experiment, triplicate determination was made and the percentage inhibition was evaluated using the mathematical expression:

Table 1FT-IR Spectra data of *V. amygdalina* column fractions.

Sample	V_{O-H} (cm^{-1})	V_{C-O} (cm^{-1})	$V_{C=O}$ (cm^{-1})	$V_{C\equiv C}$ (cm^{-1})	V_{C-H} (cm^{-1})	V_{CH} bend (cm^{-1})
Luteolin-7- α -o-glucuronide	3421	1271 1165	1710	1635	2922 2850	1377
Vernoamyoside D	3392	1263 1161	1718	1631	2937 2881	1384
Vernotolaside	3385	1263 1076		1635	2928	1411

Table 2Chemical shifts (δ) of $^1H/^{13}C$ NMR of compounds isolated from *V. amygdalina*.

Luteolin-7- α -o-glucuronide			Vernoamyoside D			Vernotolaside		
C/H Atom	^{13}C (ppm)	1H (ppm)	C/H Atom	^{13}C (ppm)	1H (ppm)	C/H Atom	^{13}C (ppm)	1H (ppm)
2	171.1	—	1	35.2	1.2(d)	1	102.31	3.40
3	104.0	6.4(s)	2	28.3	2.4(d)	2	63.31	1.07
4	172.9	—	3	72.4	3.5(s)	3	82.08	—
5	160.2	8.1(s)	4	34.5	5.3(s)	4	97.16	—
6	99.5	6.2 (brs)	5	38.6	1.6(s)	5	60.66	2.42
7	170.2	—	6	29.7	3.4(d)	6	70.65	3.70
8	90.3	6.2 (brs)	7	57.3	7.3(s)	7	56.70	—
9	152.4	—	8	60.4	—	8	75.60	0.97
10	99.6	—	9	80.3	—	9	56.70	0.85
1'	120.2	—	10	62.4	—	10	78.42	3.68
2'	110.3	7.3(S)	11	40.3	3.4	sugar		
3'	145.6	8.2 (brs)	12	36.2	5.5(d)	1'	48.13	3.40
4'	150.0	7.4 (brs)	13	39.1	1.9(d)	2'	38.40	3.20
5'	115.9	6.8(s)	14	38.3	2.1(m)	3'	40.31	3.37
6'	119.2	7.4(s)	15	56.2	2.5	4'	36.76	3.10
Sugar			16	56.3	1.9	5'	39.50	3.38
1''	99.0	5.2(s)	17	44.2	1.4	6'	44.26	3.56
2''	70.0	5.1(s)	18	56.8	2.2	—	—	—
3''	72.0	5.3(s)	19	70.4	0.8(s)	—	—	—
4''	65.0	5.2(s)	20	76.4	0.6(s)	—	—	—
5''	63.0	5.2(s)	21	60.4	—	—	—	—
6''	nil	—	22	38.2	—	—	—	—
—	—	—	23	45.1	7.4 (brs)	—	—	—
—	—	—	24	50.4	5.2(m)	—	—	—
—	—	—	25	70.3	2.1(m)	—	—	—
—	—	—	26	40.4	0.8(d)	—	—	—
—	—	—	27	38.3	0.9(d)	—	—	—
—	—	—	28	29.4	—	—	—	—
—	—	—	29	44.0	2.0(s)	—	—	—
—	—	—	Sugar			—	—	—
—	—	—	1'	74.5	4.1(d)	—	—	—
—	—	—	2'	79.0	2.8	—	—	—
—	—	—	3'	72.0	3.1	—	—	—
—	—	—	4'	77.3	3.0	—	—	—
—	—	—	5'	64.1	3.2	—	—	—
—	—	—	6'	58.3	3.4	—	—	—

$$\% \text{ Inhibition} = 100 \times (1 - V_s/V_c)$$

Where V_s = absorbance of test sample; V_c = absorbance of control.

2.5. Data analysis

IC₅₀ - Half maximal inhibitory concentration, that is, the concentration that induced 50 % inhibition was estimated by using graphpad prism (software, inc, version 8.00), Nonlinear regression (curve fit), inhibitor against normalized response-variable slope. Significant difference of the mean of the replicate values was determined by the analysis of variance (ANOVA). Values at $p < 0.05$ were considered statistically significant.

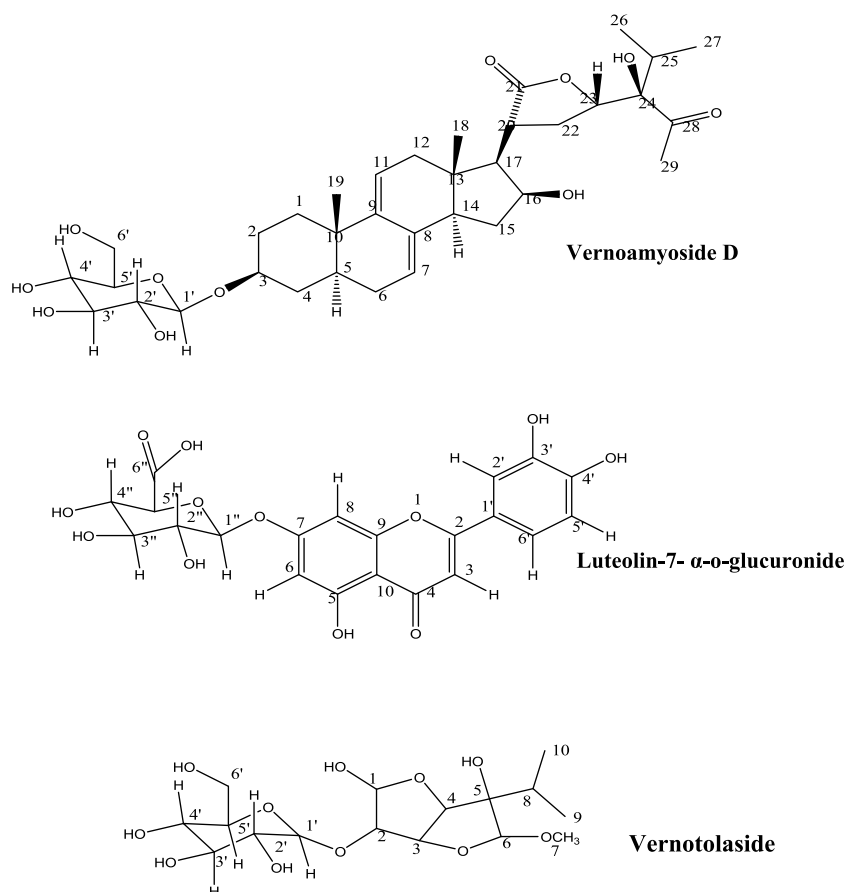


Fig. 1. Structures of Vernoaamyoside D, Luteolin-7- α -o-glucuronide and vernotolaside.

3. Results and Discussion

The gravity column chromatography of the *V. amygdalina* methanol extracts afforded two known compounds from the root and a new compound from the stem bark. The structural elucidation of the compounds was based on the combination of data obtained from the proton and carbon-13 NMR Spectroscopy, Fourier Transform Infrared (FT-IR) Spectroscopy in comparison with literature data.

3.1. Infra-red spectra of crude extracts

The IR spectra data of the isolated compounds is as indicated in [Table 1](#).

3.2. IR spectrum of luteolin-7- α -o-glucuronide

The luteolin-7- α -o-glucuronide IR spectrum exhibited stretching vibration at 3421 cm^{-1} which underscore the presence of O–H stretch of alkanols while the C=O stretching at 1710 cm^{-1} was attributed to the presence of a saturated six-membered ring ketone. The C=C absorption band at 1521 cm^{-1} was alluded to that of non-conjugated alkane while the stretching vibration at 1635 cm^{-1} was attributed to conjugated double bonds. The C–H stretching at $2922/2850\text{ cm}^{-1}$ was characteristic of the aliphatic alkane while the C–O absorption band at $1271/1165\text{ cm}^{-1}$ indicated the presence of carboxylate. The FT-IR data obtained was in agreement with literature [\[25\]](#).

3.3. IR spectrum of vernoaamyoside D

The IR spectrum of vernoaamyoside D, which was compared with literature data [\[26\]](#) exhibited characteristic O–H stretching of alcohol at 3392 cm^{-1} . The C=C absorption band appearing at 1631 cm^{-1} was attributed to the presence of conjugated alkene (Aromatic) while the vibration stretching at 1521 cm^{-1} was characteristic of an non-conjugated alkene. The C=O stretching at 1718 cm^{-1} was assigned to ketone group. The C–O stretch at 1263 cm^{-1} indicated the presence of carboxylic acid. The C–H stretch present at 2937 cm^{-1} confirmed the presence of an alkane C–H while the C–H bend of alkyl appeared at 1384 cm^{-1} .

Table 3*In vitro* anti-inflammatory (albumin denaturing potential) of crude extracts.

Conc ($\mu\text{g/mL}$)	VASAQ	VARAQ	VARMEOH	VASMEOH	VARNHEX	VASNHEX	VASETOAC	VARETOAC	Diclofenac
50	5.78 $\pm$ 4.31	10.69 $\pm$ 3.77	6.93 $\pm$ 2.75	10.11 $\pm$ 4.75	45.95 $\pm$ 4.16	43.64 $\pm$ 1.32	42.19 $\pm$ 1.52	37.57 $\pm$ 1.00	^a 32.37 $\pm$ 1.50
100	10.11 $\pm$ 5.62	12.13 $\pm$ 4.25	14.16 $\pm$ 2.64	17.63 $\pm$ 1.80	48.55 $\pm$ 4.07	48.84 $\pm$ 1.04	43.93 $\pm$ 1.04	43.64 $\pm$ 3.04	^a 37.86 $\pm$ 1.26
200	12.13 $\pm$ 6.37	14.16 $\pm$ 2.00	17.63 $\pm$ 2.18	20.52 $\pm$ 2.52	50.00 $\pm$ 4.19	48.26 $\pm$ 0.76	45.37 $\pm$ 1.32	46.24 $\pm$ 3.12	^a 44.50 $\pm$ 0.86
300	19.94 $\pm$ 4.81	17.91 $\pm$ 0.57	21.67 $\pm$ 1.60	22.83 $\pm$ 2.00	50.28 $\pm$ 3.62	50.86 $\pm$ 0.57	53.17 $\pm$ 1.32	44.79 $\pm$ 2.08	^a 52.60 $\pm$ 4.73
400	28.61 $\pm$ 1.60	19.94 $\pm$ 0.76	28.32 $\pm$ 0.76	25.14 $\pm$ 2.31	52.89 $\pm$ 3.21	54.62 $\pm$ 1.15	54.62 $\pm$ 4.62	51.15 $\pm$ 1.75	^a 64.74 $\pm$ 0.76
IC ₅₀ $\pm$ SEM	1092.00 $\pm$ 4.10	17729.00 $\pm$ 2.04	1614.00 $\pm$ 2.18	4378.00 $\pm$ 2.00	192.90 $\pm$ 4.19	195.50 $\pm$ 0.57	233.80 $\pm$ 1.34	446.30 $\pm$ 2.08	214.40 $\pm$ 0.88

Expressed data are mean $\pm$ standard error of mean of determinations made in triplicate.VASAQ – *V. amygdalina* aqueous extract (stem), VARAQ – *V. amygdalina* aqueous extract (root), VARMEOH – *V. amygdalina* methanol extract (root), VASMEOH – *V. amygdalina* methanol extract (stem), VARAHX – *V. amygdalina* n-hexane extract (root), VASEHEX – *V. amygdalina* n-hexane extract (stem), VASETOAC – *V. amygdalina* ethylacetate extract (stem), VARETOAC – *V. amygdalina* ethylacetate extract (root).^a - values at $p < 0.05$ are statistically significant.

3.4. IR spectrum of vernotolaside

The IR spectrum of vernotolaside which was compared with some compounds in literature [26] confirms the presence of O–H of alcohol at 3385 cm^{-1} while the characteristic C–O stretching at 1263 cm^{-1} indicates the presence of carboxylate. The C=C absorption band at 1635 cm^{-1} corresponded to that of alkenes (aromatic) and the alkane C–H stretch was confirmed at 2928 cm^{-1} . The C–H bend of alkene was exhibited at 1411 cm^{-1} .

3.5. NMR data

The chemical shifts (δ) of the purified isolates, vernoa-myoside D, luteolin-7- α -o-glucuronide, and vernotolaside obtained from methanol extract of *V. amygdalina* stem and root are depicted in Table 2.

Luteolin-7- α -o-glucuronide: Luteolin-7- α -o-glucuronide (Fig. 1) obtained as white powder was characterized using the infrared, proton and carbon-13 NMR data which were compared with the literatures [27]. The bands, 3392, 1718, 1631 and 1263 cm^{-1} corresponding to the O–H (Alcohol), C=O (carbonyl of esters), C=C_{ar} (conjugated alkene) and C–O (carboxylic acid) were confirmed. The carbonyl carbon at 171.1 ppm affirms the attribution. The compound was thus identified as luteolin 7- α -o-glucuronide (The glycone constituent was established with the presence of the singlet protons at 5.2, 5.1, 5.3, 5.2, 5.2, 1.00 ppm and the prominent –OH proton signals resonating at 4.70–5.01 ppm). Luteolin-7-glucuronide is a known compound that has been confirmed in various medicinal plants and herbs. Its present has been confirmed in the aerial part *Antirrhinum majus* where it was confirmed to exhibit strong antimicrobial potential [28]. It has also been isolated in *Plantago lanceolata* [29], *Salvia ekimiana* Celep & Doğan [30], *Lycopus europaeus* L. [31], *Crataegus x macrocarpa* [32] and *Perilla frutescens* (L.) Britt [33].

Vernoa-myoside D: Following the extensive evaluation of the NMR data of the compound and in comparison with literature report [34], the structure (Fig. 1) was established without ambiguity. The glycosidic character of the compounds was affirmed by the proton and carbon signals at δ 5.45 and 74.5 respectively. This position was corroborated by the presence of the OH band at 3421 cm^{-1} in the infrared spectrum. Consequently, the identification of the compound as Vernoa-myoside D was evidently aided by the combination of NMR data and the infrared report.

Vernoa-myoside D, a stigmastane-type steroidal saponin has been isolated from the *V. amygdalina* of Chinese origin previously [34]. It was shown to possess varying degree of cytotoxicity on various cancer cell line. Vernonioside B₂ with vernoniamyoside A–D obtained from the leave of the plant alongside vernoa-myoside D reportedly exhibited cytotoxicity on epithelial cells [34]. Other triterpenoids (stigmastane-type steroidal saponins; vernoa-myosides and vernoniosides have been isolated from *Vernonia britteniana* also of the Asteraceae family. The compounds exhibited high *in vitro* cercaricidal activity with moderate *in vitro* antioxidant activities [35].

Vernotolaside: Vernotolaside, a quasi-new compound had structural similarity with bifuran glycoside. The NMR data (Table 2) structure was equivocal, as no additional 2D-NMR spectroscopic data was available. The total characterization will be done when the complete spectroscopic data are obtained.

3.6. In vitro albumin denaturing assay result

The *in vitro* albumin denaturation inhibition assay result for the crude extracts: VASAQ, VARAQ, VARMEOH, VASMEOH, VARNHEX, VASNHEX, VASETOAC and VARETOAC in comparison with diclofenac (control drug) are as presented, Table 3.

The process by which external stimuli like heat, acid, base, organic chemicals, or inorganic salts trigger the protein to denature its tertiary and secondary structures is recognized as denaturation of protein [36,37] (Leelaprakash and Dass, 2011; SSen et al., 2015).

Table 4
Albumin denaturation inhibition potential of *V. amygdalina* isolated compounds.

Conc. (µg/mL)	Luteolin-7- α -o-glucuronide	Vernoamyoside D	Vernotolaside	Diclofenac
50	^a 10.19 $\pm$ 1.49	^a 17.84 $\pm$ 1.57	^a 34.56 $\pm$ 3.21	36.82 $\pm$ 0.56
100	^a 23.79 $\pm$ 1.49	^a 30.59 $\pm$ 1.49	45.60 $\pm$ 5.12	45.60 $\pm$ 0.98
200	^a 33.71 $\pm$ 0.84	^a 36.54 $\pm$ 1.41	^a 47.30 $\pm$ 3.89	52.97 $\pm$ 0.28
300	^a 47.02 $\pm$ 1.57	^a 41.07 $\pm$ 0.28	^a 55.24 $\pm$ 0.75	56.09 $\pm$ 1.13
400	^a 49.29 $\pm$ 1.13	^a 44.75 $\pm$ 1.47	^a 56.94 $\pm$ 0.28	57.50 $\pm$ 0.49
IC ₅₀ $\pm$ SEM (µg/mL)	379.5 $\pm$ 1.49	549.8 $\pm$ 1.47	201.7 $\pm$ 3.89	167.8 $\pm$ 0.28

Data are expressed as mean $\pm$ standard error of mean of triplicate determinations.

^a - values at $p < 0.05$ are statistically significant.

Nonsteroidal anti-inflammatory drugs (NSAIDs) which acts as antigens has been used for the prevention of the protein denaturation [38]. However, the several side effects which included the induction of gastric irritation leading to gastric ulcers make the option less desired [39]. Hence, plant extracts or plant-derived compounds have reported some successes as protein denaturing inhibitions [40].

The VARNHEX and VASNHEX had higher albumin denaturing inhibition with IC₅₀ at 192.9 and 195.5 µg/mL respectively than the polar extracts and the standard drug, diclofenac with IC₅₀ 214.4 µg/mL. The strength of other extracts in the anti-inflammatory activity in decreasing order is thus: VASETOAC, VARETOAC, VASAQ, VARMEOH, VASMEOH and VARAQ. These activities were however lower when compared to VARNHEX, VASNHEX and diclofenac. Hence, VARNHEX and VASNHEX exhibited more potency as an anti-inflammatory substance.

The isolated compounds recorded moderate half-maximal inhibitory concentrations (IC₅₀) thus: luteolin-7- α -o-glucuronide (379.5 µg/mL), vernoamyoside D (549.8 µg/mL) and vernotolaside (201.7 µg/mL) (Table 4). The IC₅₀ values obtained were lower than that of diclofenac, 167.8 µg/mL. These compounds showed dose-dependent inhibition activities.

The test samples showed significant dose-response albumin denaturation in the order; diclofenac, vernotolaside, luteolin-7- α -o-glucuronide and vernoamyoside D. The vernotolaside showed activity that is significantly comparable to the control, diclofenac. The result affirms that the *V. amygdalina* contain compounds with significant albumin denaturing inhibition potential. Plants are known to possess important bioactives with therapeutic potentials that are often useful for various applications particularly when preserved in appropriate form from the source [41]. Following the availability of essential minerals and bioactive compounds, folklores or plant-based formulations and decoctions are widely known for their ability to attenuate series of biological processes [42–44]. A polyherbal mixture containing *V. amygdalina* extract reportedly exhibited hepato and nephron-protective effect in animal model [45].

4. Conclusion

In this study, bioactive compounds with potential albumin denaturing inhibition potential were isolated using silica gel column chromatography from the *V. amygdalina* root and stem-bark. Utilising data from infrared, 1H, and 13C nuclear magnetic resonance spectroscopies as well as standard literatures, the characterization and structural elucidation of the purified compounds were achieved. The *in vitro* albumin denaturing inhibition was carried out on the extracts and isolated compounds using diclofenac as a standard. Three compounds which includes: luteolin-7- α -o-glucuronide, vernoamyoside D and vernotolaside were obtained. The compounds exhibited dose-dependent albumin denaturation inhibition with IC₅₀ values of 549.8 µg/mL, 379.5 µg/mL and 201.7 µg/mL respectively. The vernotolaside would require further spectroscopic characterizations and *in vivo* biological evaluations. Apparently, *V. amygdalina* holds promise as a renewable source of bioactive natural products with prospective applications in the nutritional, pharmaceutical, and nutraceutical sectors. The findings of this study lend some credibility to the claim about the traditional application of the plant in alleviating issues associated with inflammation. However, further *in vivo* studies are required for the validation of the potentials of the isolated bioactives.

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CRedit authorship contribution statement

Olubunmi Atolani: Writing – review & editing, Supervision, Software, Resources, Project administration, Methodology, Investigation, Funding acquisition, Formal analysis, Data curation, Conceptualization. **Mohammed Abubakar Usman:** Writing – original draft, Resources, Project administration, Methodology. **Jamiu Opeyemi Adejumo:** Methodology. **Adedamola Elizabeth Ayeni:** Resources, Methodology, Formal analysis. **Olamilekan Joseph Ibukun:** Validation, Formal analysis. **Adeola T. Kola-Mustapha:** Project administration, Funding acquisition. **Ngaitad S. Njinga:** Resources, Project administration, Funding acquisition. **Luqman A. Quadri:** Resources, Project administration, Funding acquisition. **Emmanuel O. Ajani:** Project administration, Funding acquisition. **Tajudeen O. Amusa:** Project administration, Funding acquisition. **Moji T. Bakare-Odunola:** Validation, Project administration, Funding acquisition. **Adenike T. Oladiji:** Resources, Project administration, Funding acquisition. **Athba Alqahtani:** Funding acquisition, Resources, Validation, Writing – review & editing. **Mohamed Abbas:** Funding acquisition, Resources, Visualization, Writing – review & editing. **Learnmore Kambizi:** Resources, Funding acquisition.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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References

- [1] Oluwaseun Ruth Alara, Hamid Abdurahman Nour, Ishmael Ukaegbu Chinonso, Ahmed Kabbashi Nassereldeen, Extraction and characterization of bioactive compounds in *Vernonia amygdalina* leaf ethanolic extract comparing Soxhlet and microwave assisted extraction techniques, *J. Taibah Univ. Sci.* 13 (1) (2019) 414–422, <https://doi.org/10.1080/16583655.2019.1582460>.
- [2] A.B. Nursuhaili, A.S.P. Nur, M.Y. Martini, M. Azizah, T.M.M. Mahmud, Medicinal values, agronomic practices and postharvest handlings of *Vernonia amygdalina*, *Food Res.* (2019) (2019).
- [3] I.I. Ijeh, C.E.C.C. Ejike, Current perspectives on the medicinal potentials of *vernonia amygdalina* Del., *J. Med. Plants Res.* 5 (7) (2011) 1051–1061.
- [4] S.A. Audu, A.E. Taiwo, A.R. Ojuolapa, A.S. Sani, A.R. Bukola, I. Mohammed, A study review of documented phytochemistry of *Vernonia amygdalina* as the basis for pharmacological activity of plant extract, *Journal of Natural Science Resources* 2 (7) (2012) 2224–3186.
- [5] T. Abiola, E.O. John, I.T. Sossou, B. Charles Callistus, Immune boosting and ameliorative properties of aqueous extract of *Vernonia amygdalina* Delile against MSG-induced genotoxicity: an *in silico* and *in vivo* approach, *Heliyon* 10 (1) (2023 Dec 3) e23226.
- [6] W.Y. Ho, S. Yeap, W.S. Liang, B.K. Beh, N. Mohamad, N.B. Alitheen, In vitro antioxidant and in vivo hepatoprotective effect on ethanol-mediated liver damage of spray dried *Vernonia amygdalina* water extract, *Pak. J. Pharm. Sci.* 28 (1) (2015) 15–22.
- [7] R.A. Syahputra, U. Harahap, A. Dalimunthe, M. Pandapotan, D. Satria, Protective effect of *Vernonia amygdalina* Delile against doxorubicin-induced cardiotoxicity, *Heliyon* 29 (7) (2021) e07434, 7.
- [8] R.A. Syahputra, U. Harahap, Y. Harahap, A.P. Gani, A. Dalimunthe, A. Ahmed, S. Zainalabidin, *Vernonia amygdalina* ethanol extract Protects against doxorubicin-induced cardiotoxicity via TGF β , Cytochrome c, and apoptosis, *Molecules* 28 (11) (2023 24) 4305.
- [9] D.B. Asante, I.T. Henneh, D.O. Acheampong, F. Kyei, C.K. Adokoh, E.G. Ofori, N.K. Domey, E. Adakudugu, L.P. Tangella, E.O. Ameyaw, Anti-inflammatory, antinociceptive and antipyretic activity of young and old leaves of *Vernonia amygdalina*, *Biomed. Pharmacother.* 111 (2019) 1187–1203.
- [10] O.J. Olorotimi, F.A. Gbore, O.D. Oloruntimehin, O.A. Jimoh, Serum inflammation and oxidative DNA damage amelioration in cocks-fed supplemental *Vernonia amygdalina* and zinc in aflatoxin B₁ contaminated diets, *Transl Anim Sci* 7 (1) (2023 16) txad113.
- [11] W.T. Wang, S.F. Liao, Z.L. Wu, C.W. Chang, J.Y. Wu, Simultaneous study of antioxidant activity, DNA protection and anti-inflammatory effect of *Vernonia amygdalina* leaves extracts, *PLoS One* 15 (7) (2020 13) e0235717.
- [12] P.A.Z. Hasibuan, J.M. Keliat, M.F. Lubis, A. Nasution, The ethyl acetate extract of *Vernonia amygdalina* leaf ameliorates gemcitabine effect against migration and invasion of PANC-1 cells via down-regulation of the VEGF, COX₂, and RAS/MEK pathways, *Saudi Pharmaceut. J.* 32 (1) (2024) 101872.
- [13] M. Tesfaye, Y. Gonfa, G. Tadesse, T. Temesgen, S. Periyasamy, Green synthesis of silver nanoparticles using *Vernonia amygdalina* plant extract and its antimicrobial activities, *Heliyon* 15 (9) (2023) e17356, 6.
- [14] C.B. Adeosun, S. Sinmisola, A.O. Opeifa, O.*. Atolani, Essential oil from the stem bark of *Cordia sebestena* scavenged free radicals, *Journal of Acute Medicine* 3 (2013) 138–141.
- [15] O. Atolani, O.S. Adeyemi, E. Akpan, C.B. Adeosun, G.A. Olatunji, Chemical composition and antioxidant potentials of *Kigelia pinnata* root oil and extracts, *EXCLI Journal* 10 (2011) 264–273.
- [16] O. Atolani, A. Adeniji, O.O. Omonike, G.A. Olatunji, Energy dispersive - XRF metal analysis and cancer cell line cytotoxicity of *Kigelia pinnata* root, *Journal of Biologically Active Products from Nature* 3 (3) (2013) 194–199.
- [17] O. Atolani, G.A. Olatunji, O.S. Adeyemi, Cytotoxicity of lapachol and derivatized analogues from *Kigelia africana* (lam.) benth. On cancer cell lines, *Arabian J. Sci. Eng.* 46 (2021) 5307–5312.
- [18] G.N. Elemo, O.L. Erukainure, J.N.C. Okafor, P. Banerjee, R. Preissner, Nwachukwu, A. Viola, O. Atolani, A. Awosika, F. Shode, Underutilized legumes, *Cajanus cajan* and *Glycine max* may bring about antisickling effect in sickle cell disease by modulation of redox homeostasis in sickled erythrocytes and alteration of its functional chemistry, *J. Food Biochem.* 46 (9) (2022) e14322.
- [19] Z. Wang, R. Ma, B. Chen, X. Yu, X. Wang, X. Zuo, J. Yang, A transcription factor-based bacterial biosensor system and its application for on-site detection of explosives, *Biosens. Bioelectron.* 244 (2024) 115805.
- [20] M. Li, S. Lv, R. Yang, X. Chu, X. Wang, Z. Wang, J. Yang, Development of lycopene-based whole-cell biosensors for the visual detection of trace explosives and heavy metals, *Anal. Chim. Acta* 1283 (2023) 341934.
- [21] H. Liu, P. Kang, Y. Liu, Y. An, Y. Hu, X. Jin, X. Cao, Y. Qi, T. Ramesh, X. Wang, Zinc oxide nanoparticles synthesised from the *Vernonia amygdalina* shows the anti-inflammatory and antinociceptive activities in the mice model, *Artif. Cells, Nanomed. Biotechnol.* 48 (1) (2020) 1068–1078.
- [22] R. Zhang, X. Chen, Y. Cheng, Z. Chen, X. Li, Y. Deng, Recent advances of nanomaterials for intervention in Parkinson's disease in the context of anti-inflammation, *Coord. Chem. Rev.* 502 (2024) 215616.
- [23] O. Atolani, O.E. Olorundare, A.N. Anoka, A.O. Osin, S.A. Biliaminu, Antioxidant, proteinase inhibitory and membrane stabilization potentials of *Moringa oleifera* seed oil, *Fabid J. Pharm. Sci.* 43 (2) (2018) 1–13.
- [24] L. Kambizi, M.T. Bakare-Odunola, A.T. Oladiji, A.T. Kola-Mustapha, T.O. Amusa, O. Atolani, N.S. Njinga, A.L. Quadri, Protease inhibition, membrane stabilization, antioxidant and phytochemical evaluations of leaves, seeds and calyces of four selected edible medicinal plants, *Cogent Chemistry* 3 (2017) 1314064.
- [25] Hao Dong, Xiaocui Yang, Jiapeng He, Sheng Cai, Kaijun Xiao, Liang Zhu, Enhanced antioxidant activity, antibacterial activity and hypoglycemic effect of luteolin by complexation with manganese (ii) and its inhibition kinetics on xanthine oxidase, *RSC Adv.* 7 (84) (2017) 53385–53395.
- [26] Meng-Ling Zhao, Shu-Jun Shan, Rong Tao, Le-Tian Cui, Qiu-Rong Li, Jun Luo, Li Yi, Stigmastane-type steroid saponins from the leaves of *Vernonia amygdalina* Del., *Fitoterapia* 150 (104838) (2021) 1–8.
- [27] D. Ryu, H.J. Jee, S.Y. Kim, S.H. Hwang, G.B. Pil, Y.S. Jung, Luteolin-7-O-Glucuronide improves depression-like and stress coping behaviors in sleep deprivation stress model by activation of the BDNF signaling, *Nutrients* 12 (14) (2022) 3314, 16.
- [28] F.G. Saqallah, W.M. Hamed, W.H. Talib, R. Dianita, H.A. Wahab, Antimicrobial activity and molecular docking screening of bioactive components of *Antirrhinum majus* (snapdragon) aerial parts, *Heliyon* (8) (2022 27) e10391, 8.
- [29] L.H. Bajrai, A.S. Alharbi, M.M. El-Day, A.G. Bafaraj, V.D. Dwivedi, E.I. Azhar, Identification of antiviral compounds against monkeypox virus profilin-like protein A42R from *Plantago lanceolata*, *Molecules* 27 (22) (2022 9) 7718.

- [30] G.Ş. Karatoprak, F. Göger, İ. Çelik, Ü. Budak, E.K. Akkol, M. Aschner, Phytochemical profile, antioxidant, antiproliferative, and enzyme inhibition-docking analyses of *Salvia ekimiana* Celep & Doğan, S Afr J Bot. 146 (2022 May) 36–47, <https://doi.org/10.1016/j.sajb.2021.09.033>.
- [31] H.G. Gumbinger, H. Winterhoff, R. Wylde, A. Sosa, On the influence of the sugar moiety on the antigonadotropic activity of luteoline glycosides, Planta Med. 58 (1) (1992) 49–50.
- [32] A. Ringl, S. Prinz, A. Huefner, M. Kurzmann, B. Kopp, Chemosystematic value of flavonoids from *Crataegus x macrocarpa* (Rosaceae) with special emphasis on (R)- and (S)-eriodictyol-7-O-glucuronide and luteolin-7-O-glucuronide, Chem. Biodivers. 4 (2) (2007 Feb) 154–162.
- [33] Z. Wu, S. Lee, B. Kang, S. Lee, K. Koo, J. Lee, S. Lim, Determination of luteolin 7-glucuronide in *Perilla frutescens* (L.) Britt. Leaf extracts from different regions of China and Republic of Korea and its cholesterol-lowering effect, Molecules 28 (20) (2023 Oct 10) 7007.
- [34] J. Wang, H. Song, X. Wu, S. Zhang, X. Gao, F. Li, X. Zhu, Q. Chen, Steroidal saponins from *vernonia amygdalina* del. And their biological activity, Molecules 23 (3) (2018. 5) 579.
- [35] M.D.A. Valente, P. Ferreira, K. Lima, I.B. Moreira da Silva, P. Nobre, I. Neto, M. Pires, B.S. Braz, R. Serrano, S. Belo, O. Silva, *Vernonia britteniana* root phytochemical studies, in vitro cercaricidal activity on the larval stage of *Schistosoma mansoni* and antioxidant activities, Plants 12 (9) (2023 27) 1788.
- [36] G. Leelaprakash, S.M. Dass, In vitro anti-inflammatory activity of methanol extract of *Enicostemma axillare*, J Drug Dev Res. 3 (2011) 189–196.
- [37] S. Sen, R. Chakraborty, N. Maramsa, M. Basak, S. Deka, et al., In vitro anti-inflammatory activity of *Amaranthus caudatus* L leaves, Indian J Nat Prod Resour 6 (2015) 326–329.
- [38] P.A. Insel, Analgesic-antipyretic and anti-inflammatory agents and drugs employed in the treatment of gout, in: J.G. Hardman, L.E. Limbird, P.B. Molinoff, R. W. Ruddon, A. Gilman (Eds.), The Pharmacological Basics of Therapeutics, ninth ed., McGraw Hill, New York, 1996, pp. 617–657.
- [39] M. Marliyah, T. Ananthi, In vitro anti-inflammatory activity of extract of *Zea mays* (L.), J Glob Biosci. 4 (2015) 2168–2173.
- [40] S. Dharmadeva, L.S. Galgamuwa, C. Prasadinie, N. Kumarasinghe, In vitro anti-inflammatory activity of *Ficus racemosa* L. bark using albumin denaturation method, Ayu 39 (4) (2018) 239–242.
- [41] J. Xiang, R. Mlambo, I. Shaw, Y. Seid, H. Shah, Y. He, B. He, Cryopreservation of bioflavonoid-rich plant sources and bioflavonoid-microcapsules: emerging technologies for preserving bioactivity and enhancing nutraceutical applications, Front. Nutr. 10 (2023), <https://doi.org/10.3389/fnut.2023.1232129>.
- [42] K. Parajuli-Baral, Formulation and evaluation of quality parameters of effervescent granules from the potent antioxidant between two variants of the adaptogenic herb *Ocimum tenuiflorum* L, Sci. World J. 2023 (2023) 2050846.
- [43] M. He, T. Ren, Z.D. Jin, L. Deng, H. Liu, Y.Y. Cheng, H. Chang, Precise analysis of potassium isotopic composition in plant materials by multi-collector inductively coupled plasma mass spectrometry, Spectrochim. Acta B Atom Spectrosc. (2023) 106781, <https://doi.org/10.1016/j.sab.2023.106781>.
- [44] B. Huang, M. Gui, H. An, J. Shen, F. Ye, Z. Ni, J. Lin, Babao Dan alleviates gut immune and microbiota disorders while impacting the TLR4/MyD88/NF-κB pathway to attenuate 5-Fluorouracil-induced intestinal injury, Biomed. Pharmacother. 166 (2023) 115387.
- [45] O.O. Iroanya, O.A. Adebisin, J. Okpuzor, Evaluation of the hepato and nephron-protective effect of a polyherbal mixture using wistar albino rats, J. Clin. Diagn. Res. 8 (6) (2014) HC15–21.